



Extraction and modification of cellulose from peanut shells and cornstalks and for use as adsorbents for removal of lead

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ABSTRACT

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The requirement for novel adsorbents from inexpensive sources, directs this research towards innovation of new adsorbent materials through extracting and modifying cellulose fibres from peanut shells and corn stalk. The processes involved delignification, alkaline hydrolysis, acetylation of pure cellulose and the adsorption of Lead Pb^{2+} . The adsorbents obtained are pure celluloses of peanut shells (PSC), Corn stalk (CSC) and Acetyled Celluloses of Peanut Shells (PSCA) and Corn Stalk (CSCA). The Structural and functional properties were analyzed by Fourier transform infrared spectroscopy (FTIR) and Atomic Absorption Spectrometer (AAS). The four adsorbents all exhibited high removal percentage of Lead from the solutions. However, acetyled Cellulose of peanut shells exhibited the highest adsorption capacity of 397.5 mg/g with final Lead (Pb) concentration of 0.125 mg/L, the removal of Lead from solution amounted to 99.3% as compared to the other adsorbents. This research proved the efficiency of agro-waste cellulose acetate for use as novel adsorbents through the Lead Adsorption. An extensive exploration in researches involving biodegradable waste materials is required to utilize this source and control environmental pollution.

1. Introduction

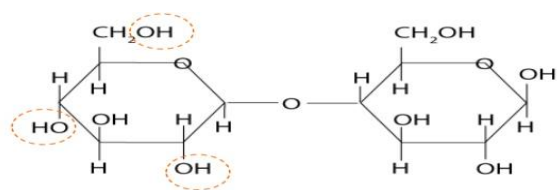
It has become a challenging quest to replace toxic and non-renewable petrochemical products in the world with hopes of reducing contamination of the Ecosystems. These Ecological concerns have resulted in a renewed interest in natural, renewable resources- based and compostable materials. For these reasons, material components such as natural fibers, biodegradable polymers can be considered 'environmentally safe' alternatives for the development of new biodegradable adsorbents, which can be used as media for pollution control [1]. It is of utmost relevance that environmental pollution in affected parts of Nigeria and other developing states should be tackled with in order to ensure healthy progression of these nations. The purification of water and other systems against heavy metals has long been under study using low-cost adsorbents from plant wastes. Bioremediation and other

conventional methods of purification have been implored but have not yet proven satisfactory due to some technical and economical constraints.

Cellulose is one of the most widespread biopolymer found globally, existing in a variety of living species such as plant, bacteria and some amoebas [2]. Cellulose is the primary component of the cell walls of higher plants. It comprises at least one third of the vegetable matter of the world [3].

Natural fibers mainly consist of cellulose, lignin, and hemicellulose but also include low quantities of pectin, pigments and extracts. The natural fibers themselves act as composite materials, assembling in a mainly lignin matrix [4]. These cellulosic materials are well known for their abundance, economic and environmental advantages.

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Structure 1. Structure of Cellulose

The availability of OH groups facilitates the degree of Hydrogen-bonding in cellulose. The degree of substitution is the term used to express the average number of hydroxyl group replaced [5]. The Drawbacks of cellulose are the high number of hydroxyl groups, which lead to strong hydrogen interactions between two fibers and to the gel-like structure once produced. The second drawback is the high hydrophilicity of the material, which limits its uses in several applications such as in paper coating (increase of dewatering effect) or composites (tendency to form agglomerates in petrochemical polymers [6]. However, surface modification with addition of substances such as silica SiO_2 have been discovered to reduce hydrophilicity of cellulose composites [7].

This investigation purports to discover the high adsorption capacity of extracted cellulose before after its modification to cellulose acetates. Peanut shells and corn stalks are abundant and inexpensive agricultural waste products that have relatively high cellulose content of 65.5 - 79.3% [8]. Nigeria is the third largest peanut producer in world, having a record of producing 2,755,649 metric tons annually, according to WorldAtlas (2016). Nutritional analysts found Peanut shells to be composed of 65.7% cellulose, 21.2% carbohydrates, 7.3% protein, 4.5% minerals and 1.2% lipids [9] Chemical content of corn stalk contains cellulose (35-50%), lignin (5-34%) and petosan around 20-41% [10]. The utilization of peanut shells and corn Stalks is of substantial benefit and at low costs due to its abundance in the country. Their richness in cellulose provides its aptness to serve as raw material for cellulose modification[11].

In recent years, surface modifications of cellulose have been a subject of increasing interest because of their significantly enhanced mechanical properties and thermal stability compared to conventional polymer composites and adsorbents. Abundance, high Tensile strength and stiffness, low weight and biodegradability are some special useful features of cellulose fiber materials [12]. Isolation, characterization, and search for applications of novel form of cellulose, is generating much activity currently. Isolated cellulosic materials are able to combine important cellulose properties such as hydrophilicity, broad chemical-modification capacity, and the formation of versatile semi-crystalline fiber morphologies to form better adsorbents [13]. The adsorption and recovery capabilities

of cellulose acetate are being investigated for separation of precious metals such as gold and platinum from leaching processes [14].

2.Experimental

2.1 Sampling

The waste samples of peanut shells and corn stalks were obtained from a local farmhouse in Maiduguri. Samples were thoroughly milled, washed and dried to reduce waxes and debris [15].

2.2 Procedures

2.2.1 Delignification and Bleaching

A solution of 3.5% w/v NaClO_2 and 6 ml glacial acetic acid was prepared in two 1000 ml beakers. Solution is mostly active at given pH and temperature therefore a thermometer and universal litmus was used to ensure the standard pH of 3-4 and heated to 75°C . Grounded dry samples of 50 g cornstalk and peanut shells was weighed and added to heated solution. Allowed to stir gently for 1hr 30mins. The sample colour became yellow. The residue was washed thoroughly with cold water and filtered [16].

2.2.2 Alkaline Hydrolysis

This process required a solution of 18% NaOH in 800 ml of water at ambient temperature in order to dissolve the remaining hemicellulose present in delignified sample. The delignified residue was added to the solution and allowed to soak for 24 hrs [7,11]. The mixture was further neutralized with 14 ml glacial acetic acid. It was then washed and filtered repeatedly until pH became neutral[13]. The resulting product was alpha (pure) cellulose.

2.2.3 Acetylation

This process required strict measures of using a gas mask and the reaction was carried out in a fume cupboard. 100 ml glacial acetic acid was added to 20 g of each cellulose sample. 50 ml acetic anhydride and 5 ml H_2SO_4 as catalyst, was added to the mixture and stirred with a glass rod. The Reaction was at ambient temperature under a fume cupboard. Time was set for 24 hrs. 200 ml of water was added and Cellulose acetate precipitated[17]. Filtration was carried out using vacuum filter apparatus. Cellulose acetate was washed and dried at room temperature [18].

2.2.4 Adsorption of Lead Pb^{2+}

1 g of Adsorbents, Peanut shell and corn stalk cellulose and cellulose acetate were placed in four 50 ml-beaker. 20 ml standard solution of $\text{Pb}(\text{NO}_3)_2$ having Pb at 1000 mg/L of, was poured into each beaker using a measuring cylinder. The mixture was allowed to react for 15 mins and subsequently filtered out using whatman filter paper. The filtered was subjected to AAS analysis, while the residue was dried out [10,19].

2.3 Determination of percentage yields from experimental procedures

The yields of alpha cellulose from both peanut shell corn stalks were determined using equation (1.0). This formula was used to calculate the percentage yields of the extraction process of cellulose from both peanut shells and corn stalk [20,21].

$$\alpha \text{ cellulose} = \frac{\text{final weight}}{\text{initial weight}} \times 100 \text{-----(1)}$$

2.4 Atomic Adsorption Spectroscopy

The adsorption capacity Q_e is calculated using the formula:

$$Q_e = \frac{(C_i - C_f)V}{W} \text{-----(2) [11,22]}$$

The percentage removal of Pb ion from solution was calculated using the following formula:

$$\% \text{ Removal} = \frac{C_i - C_f}{C_i} \times 100 \text{-----(3) [11,23]}$$

Where Q_e is adsorption capacity, C_i is initial concentration, C_f is final concentration, V is solution volume and W is mass of adsorbent.

3. Result and discussion

3.1 Percentage Yield

The methodology of estimating percentage yield is presented in Section 2.3. The percentage yield of Corn Stalk Cellulose and Peanut Shell Cellulose was 77.1% and 78.4%, respectively. This value for peanut shells cellulose is relative with the findings of Masenda *et al* [4] whose values for cellulose from peanut shells was 65.3% - 79%. The corn stalk values were significantly higher than several researches [13, 16], whose value for sorghum stalk was 47.36%. However, the slight variations could be as result of the effect of land quality of soil that the different groundnuts were grown or experimental error.

During the Acetylation process, a fraction of approximately 20 % of the extracted Cellulose was further dissolved yielding a total of 15.5 g of Cornstalk and 15.7 g of peanut Shell cellulose acetate from 20 g of alpha cellulose. This was due to residual hemicellulose which dissolved in acetic acid [23]. Table 1. presents the yield obtained in this study.

Table 1.: Percentage yields through experimental processes

EXPERIMENTAL PROCESSES	CORN STALK			PEANUT SHELLS		
	Initial weight (g)	Final weight (g)	Percentage weight (%)	Initial weight (g)	Final weight (g)	Percentage weight (%)
Bleaching	50	31.4	62.8	50	36.1	72.2
Alkaline Hydrolysis	31.4	24.2	77.1	36.1	28.3	78.4
Acetylation	20	15.68	78.4	20	15.47	77.35

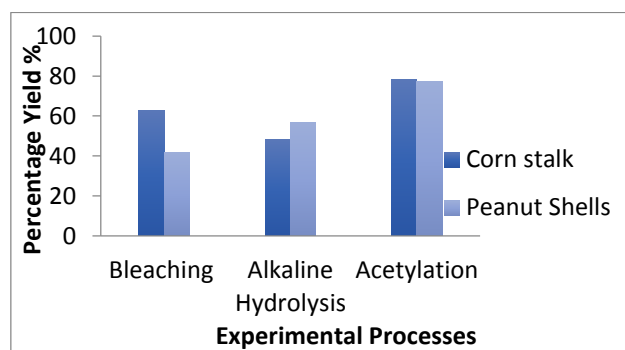


Figure 1. Percentage yields through experimental processes

3.3 FTIR Spectroscopy

The most convenient and effective method of studying the structural characteristics of cellulosic experiments is the FTIR Spectroscopy [24,28]. The following FTIR spectra Figures 2, 3, 4 and 5, depict the spectrum of Peanut Shells (PSC) and cornstalk (CSC) celluloses, Peanut Shells (PSCA) and cornstalk Cellulose Acetate (CSCA).

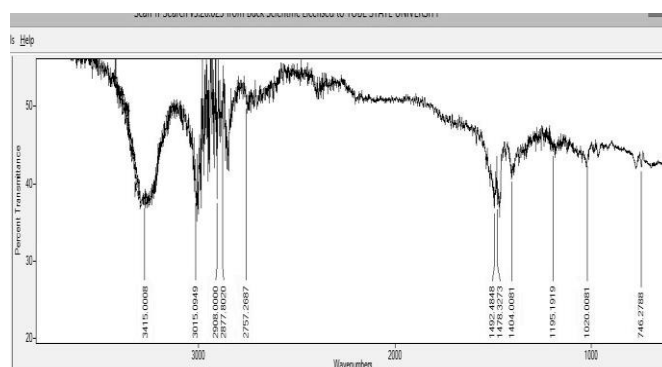


Figure 2. FTIR Spectrum of Corn Stalk Cellulose (CSC)

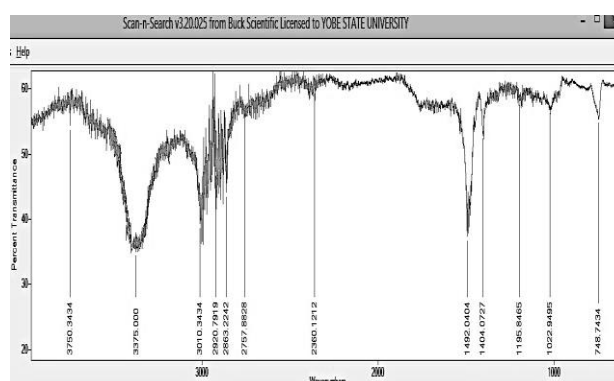
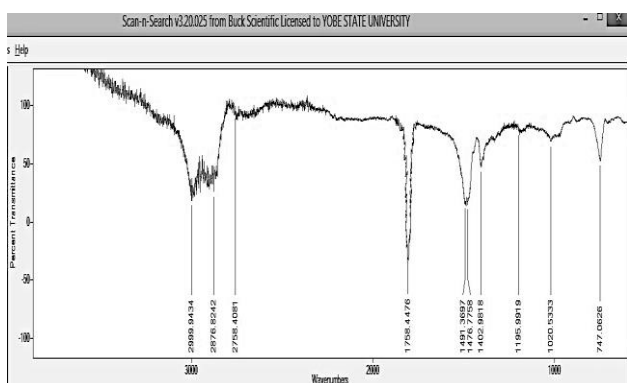
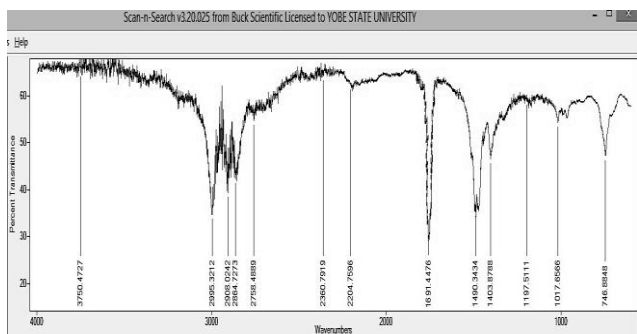


Figure 3. FTIR Spectrum of Peanut Shells Cellulose (PSC)

The spectra on figures 2 & 3 exhibit functional groups related to pure celluloses[25]. The broad band at 3415 and 3375 cm^{-1} are due to the presence of hydroxyl groups -OH. The peaks at 3010 and 1490 cm^{-1} displays -C-H stretch and bending bond vibrations[26]. The peaks at 1492 to 1404 cm^{-1} indicate C-O stretching confirming the presence of lignin[27].

Table 2.: Lead Concentrations (mg/L)

S/N	Sample Name	Sample Conc (mg/L)	Sample Conc. (mg/L)	Sample Conc. (mg/L)	Mean Conc. (mg/L)	Standard Deviation
1	PSC	0.228	0.228	0.226	0.227	0.001
2	CSC	0.228	0.224	0.231	0.228	0.004
3	CSC	0.602	0.595	0.605	0.601	0.005
4	PSC	0.126	0.122	0.128	0.125	0.003

**Figure 4.** FTIR Spectrum of Corn Stalk Cellulose Acetate (CSCA)**Figure 5.** FTIR Spectrum of Peanut Shell Cellulose Acetate (PSCA)

Both Peanut shells and corn stalk Cellulose Acetates spectra in figures 4 & 5, shows that acetylation has taken place through peaks appearing at 1758 & 1691 cm^{-1} depicting Carbonyl groups $-\text{C}=\text{O}$ and 1195 cm^{-1} for $-\text{C}-\text{O}$ stretch by acetyl groups [29]. Although the $\text{O}-\text{H}$ peaks have completely diminished in the acetyled cellulose, Peanut shells cellulose and cellulose acetate appear to have sharper peaks than modified corn stalk. This leads to the conclusion that modified peanut shells have a higher potential for more active binding sites and would

therefore be better adsorbents than the modified cornstalk[30].

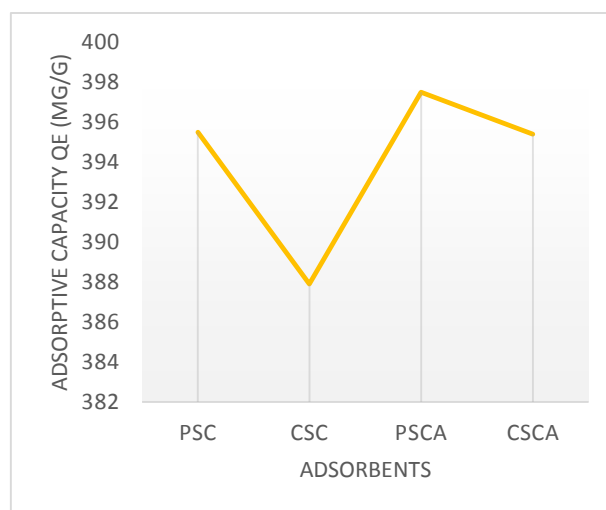
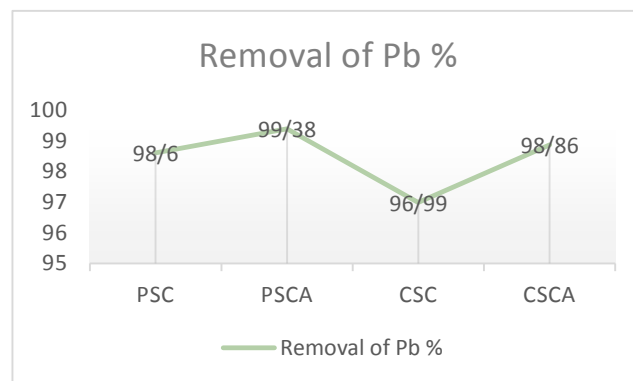
3.2. Adsorption:

3.2.1. Adsorption Capacities

The efficacy of the pure cellulose and modified cellulose samples for the adsorption of lead was determined using the AAS.

Table 3. Adsorption Isotherms

Adsorbents (g)	Pb^{2+} Conc. (mg/L)	
	Initial C_i	Final C_f
Peanut Shell-Cellulose	20	0.227
Corn Stalk-Cellulose	20	0.601
Peanut Shell-Cellulose Acetate	20	0.125
Corn Stalk-Cellulose Acetate	20	0.228

**Figure 6.** Adsorption capacity with respect to Concentrations of lead Pb^{2+} **Figure 7.** Percentage Removal of Pb^{2+} by Adsorbents

The four adsorbents all exhibited high removal percentage of Lead from the solutions as presented in the

Table 2 and Figure 6. However, surface acetylated Cellulose of peanut shells exhibited the highest adsorption capacity of 397.5 mg/g with final lead concentration of 0.125 mg/L, the removal of Lead from solution amounted to 99.3% as compared to the other adsorbents. This is as a result of the morphological and structural change of the Peanut Shells fibres through acetylation[31]. As the FTIR had shown higher peaks in the peanut shells cellulose and cellulose acetate appearing to have more active binding sites than the cornstalk cellulose acetate and non-modified cellulose[30,32].

3.2.2. Effect of adsorption on adsorbents

The dried samples of adsorbents were weighed after the adsorption process to analyse any change in weight due to deposition of Lead compound [Pb(NO₃)₂] on adsorbent surface.

Table 4.: Weight gains on Adsorbents after adsorption process

Adsorbents	Initial weight of adsorbent (g)	Final weight of adsorbent (g)	Weight gained By adsorbent (g)
Corn stalk cellulose acetate	1.0	1.06	0.06
Peanut shell Cellulose acetate	1.0	1.05	0.05
Corn stalk Cellulose	1.0	1.03	0.03
Peanut Shell cellulose	1.0	1.01	0.01

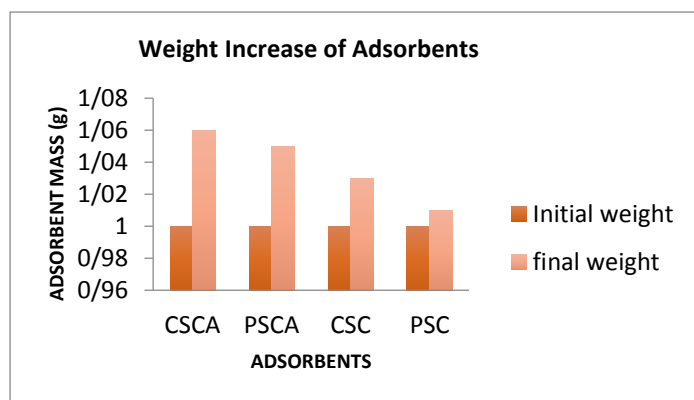


Figure 8. Weight gains on Adsorbents after adsorption process

There is an evident increase in weight of all adsorbents but a significantly higher increase in weight of the cellulose acetates which proves the theory of more active binding sites leading to higher deposition of lead (Pb) ion on the adsorbents[33]. However, PSCA should have the highest in correspondence to the results of the Removal of Pb in Figure 7, but CSCA appears to have higher weight in Table 4 and Figure 8 above. This could be as a result of error during weighing[34].

4. Conclusion

The processes of isolation of Cellulose from peanut shells and corn stalk have proven effective as it resulted to attainment of medium grade cellulose acetate, which was successfully used as an adsorbent of Lead Pb²⁺. Although, the Cellulose acetates exhibited higher absorption capacities as compared to the alpha or pure celluloses, Peanut shells cellulose acetate provided the best result for adsorptive capacity with removal of about 99.3% of Pb ions from the solution. The significant weight loss during the alkaline hydrolysis and delignification indicated that the soluble sugars amount up to half of the fibre weight. Acetylation process further dissolved 20% of the two different cellulose fibres. The quick adsorption of Pb ions by the cellulose acetates, within 10 to 15 minutes is the most remarkable breakthrough of this research. This research provides another source to be used as cheaper alternatives of adsorbents, for pollution control, as well as industrial separation of precious metals.

REFERENCES

- [1] Kurush A.S. and Sajad P (2021) Biodegradable film of black mulberry pulp pectin/chlorophyll of black mulberry leaf encapsulated with carboxymethylcellulose/silica nanoparticles: Investigation of physicochemical and antimicrobial properties. *Materials Chemistry and Physics*, 267: 124580.
- [2] Bharthare, P., Shrivastava, P., Singh, P. & Ttiwari A. (2014). Peanut shell as renewable energy source and their utility in production of ethanol. *International Journal of Advanced Research*. 2(4), 1-12
- [3] Unagolla J.M., & Adikary S.U. (2015). Adsorption Characteristics of cadmium and Lead Heavy Metals into Locally Synthesized Chitosan Bipolymer. *Tropical Agricultural Research*, 26(2), 395-401.
- [4] Masenda, E. (2004). Groundnut Shells”, *Mushroom Growers*, 5, 21-122.
- [5] Daud, Z., Hatta, M.Z.M., Kassim, A.S.M., Awang H. & Aripin, A.M. (2013). Analysis of the Chemical Composition and Fiber Morphology Structure of Corn Stalk. *Australian Journal of Basic Applied Sciences*, 7(9), 401-405.
- [6] Raabe, J., Fonesca, A., Bufalino, F., Ribeiro, C., Martins, M.A., Marconcini, J.M., Mendes, L.M. & Tonoli, G.H.D. (2015). Biocomposite of Cassava Starch Reinforced with Deposition of Silica (SiO₂) Nanoparticles. *Hindawi PC, Journal of Nanomaterials*, 1-9.
- [7] Kiro. M. (2015). “Cellulose fibers extracted from agricultural biomass.” *Scientific and Professional Journal of the Union of Textile Engineers and Technicians of Siberia*. 62(4), 15-19,
- [8] Aboody, M.H. (2013), Extraction of Cellulose from some Industrial and Plant Waste and its hydrolysis using new heterogeneous catalyst, *M.S.thesis, College of Science University of Baghdad*.1-83
- [9] Singanousong, R., Tochampa, W., Kongbangkerd, T. & Sodchit, C. (2014). Extraction and Properties of Cellulose from Banana Peels. *Suranaree Journal of Science and Technology*. 21, 201-213.
- [10] Achor, M., Oyeniyi, Y.J. & Yahaya, A. (2014). Extraction and characterization of microcrystalline cellulose obtained from the back of the fruit of Lageriana siceraria (water gourd). *Journal of Applied Pharmaceutical Science*. 4 (01), 057-060.

- [11] Sajad P., Elnaz F. and Leila R. (2020) Antioxidant/Antimicrobial Film Based on Carboxymethyl Cellulose/Gelatin/TiO₂-Ag Nano- Composite. *Journal of Polymers and the Environment*.
- [12] Arezu J., Sajad P., Mir K.P. & Saber A. (2021) Biodegradable Nanocomposite Film Based on Gluten/Silica/Calcium Chloride: Physicochemical Properties and Bioactive Compounds Extraction Capacity. *Journal of Polymers and the Environment* 29, 2557–2571
- [13] Somayeh M., Vahid R. and Sajad P. (2019) Design and fabrication of nanocomposite-based polyurethane filter for improving municipal waste water quality and removing organic pollutants Adsorption Science & Technology, 37(1–2): 95–112
- [14] Fariba G., Sajad P., Mohammad Al. & Forough M. (2017) Extraction and determination of volatile organic acid concentration in pomegranate, sour cherry and red grape juices by PPy-Ag nanocomposite fiber for authentication. *Separation Science and Technology*, <http://dx.doi.org/10.1080/01496395.2017.1380668>.
- [15] Sara K., Leila R., Sajad P. and Mehdi G. (2020) Green extraction of bioactive compounds of pomegranate peel using Cyclodextrin and ultrasound. *Main Group Chemistry*, 19: 61–80
- [16] Dimitrios L., Prodromos S. and Stefanos L. (2021) Technologies and Extraction Methods of Polyphenolic Compounds Derived from Pomegranate (*Punica granatum*) Peels A Mini Review. *Processes*, 9, 236.
- [17] Ali J., Sajad P. and Morteza B. (2017) Synthesis and characterization of magnetic nanocomposites based on Hydrogel-Fe₃O₄ and application to remove of organic dye from waste water. *Main Group Chemistry*, 16 85–94
- [18] Sajad P. and Farrokh A. (2021) Synthesis of Fe₃O₄ /SiO₂ /Polypyrrole magnetic nanocomposite polymer powder: Investigation of structural properties and ability to purify of edible sea salts. *Advanced Powder Technology*, 32(4): 1233-1246
- [19] Kamaruzzaman, S., Aris, N.I.F., Yahaya, N., Hong, L.S. & Razak, M.R., (2017). Removal of Cu and Cd Ions from Environmental water samples by using Cellulose acetate membrane. *Journal of Environmental Analytical Chemistry*. 4(4), 1-8.
- [20] Abbakar, M.A. & Elsheikh, M.T. (2015), Investigation of α -Cellulose Local Sources for the Production of Nitrocellulose, *International Journal of Sciences, Khartoum, Sudan*, 19, 35-50.
- [21] Kopania, E., Wietecha, J. & Ciechańska, D. (2006). Studies on Isolation of Cellulose Fibres from Waste Plant Biomass. *Instytut Biopolimerów and Chemical Fibres*. 20, 6B (96), 167-172.
- [22] Missoum, K., Belgacem, M.N. & Bra, J. (2013). Nanofibrillated Cellulose Surface Modification. *Material*, 6, 1745-1766.
- [23] Kontturi, E. (2015). “Cellulose: Chemical Modification”, *Lecture CHEM-E2140, School of chemical Technology, Aalto University*.
- [24] Laxmeshwar, S.S., Viveka, S., Kumar, M., D.J. & Nagaraja, G.K. (2012). Preparation and properties of composite films from modified cellulose fibre-reinforced with PLA. *Der Pharma.Chemica*. 4(1), 159-168.
- [25] Junka, K., Filpponen, I., Johansson, L.S., Kontturi, E., Rojas, O.J. & Laine, J. (2014). A method for the heterogeneous modification of nanofibrillar cellulose in aqueous media. *Carbohydrate Polymers, Elsevier*. 107–115.
- [26] Garcia, R. & Baez A. P. (2012). Atomic Adsorption Spectrometry (AAS). In *Tech. Global Acetate Manufacturers Association GAMA (2017). Cellulose Acetate Polymer*
- [27] Bouhdadi, R., Benhadi, S., Molina, S., George, B., El Moussauiti, M. & Merlin, A. (2011). Chemical Modification of Cellulose by Acetylation: Application to adsorption of methylene blue. *Maderas*.
- [28] Avérous, L. & Le Digabel, F. (2006). Properties of Biocomposites Based on Lignocellulose Fillers. *Carbohydrate Polymers, Elsevier*, 1-14 *Ciencia y Tecnologia*. 13(1), 105-116.
- [29] Brydson, J.A. (1999). *Plastic Materials- Cellulose Plastics. 7th Edition*, pp. 613-634.
- [30] Kalia, S., Dufresne, A., Cherian, B.M., Kaith, B.S., Avérous L., Njuguna J., & Nassiopoulos, E. (2011). Cellulose based biocomposites and nanocomposites. *International Journal of Polymer Science*. 1-30
- [31] Patel, S.H., & Patel, H. N. (1989). Cellulose Acetate. Interplex India Private Limited, <http://www.interplexindia.com/aca.htm>
- [32] Oлару, N., Oлару, L., Vasile, C. & Ander, P. (2011). Surface modified cellulose obtained by acetylation without solvents of bleached and unbleached kraft pulp. *Polimery*, 56, 834-840.
- [33] Zargar B., Parham H. & Shiralipour R. (2017), Removal of Pb and Cd ions from contaminated water by dithizone-modified cellulose acetate nanosponges. *Journal of Materials and Environmental Sciences*, 8(3), 1039-1045.
- [34] Yang J., Kubota F., Baba Y., Kamiya N., Goto M. (2014) “Application of cellulose acetate to the selective adsorption and recovery of Au(III)”. *Carbohydrate polymers*. Vol:111, pp: 768-774.